

Clitoria mariana var. *mariana*

Butterfly-pea

Fabaceae



Clitoria mariana var. *mariana* by Kevin Metcalf, 2021

Clitoria mariana var. *mariana* Rare Plant Profile

New Jersey Department of Environmental Protection
State Parks, Forests & Historic Sites
Forests & Natural Lands
Office of Natural Lands Management
New Jersey Natural Heritage Program

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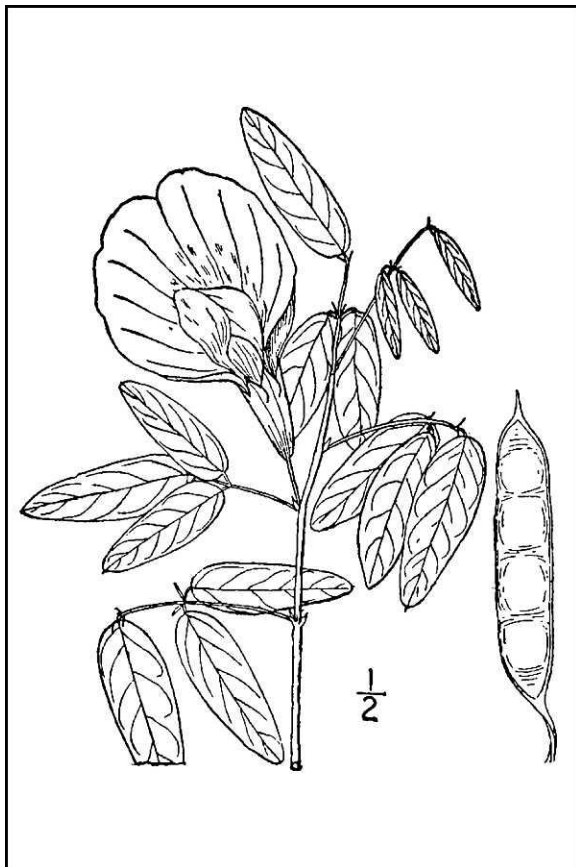
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For:
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Life History

Clitoria mariana (Butterfly-pea) is a perennial vine in the Fabaceae that was described as "a very elegant plant" by Barton (1818). The smooth stems are low and erect or twining, usually remaining under a meter in length. The leaves are alternate and divided into three stalked leaflets that are oblong to ovate and bluntly tipped. Narrow stipules are present at the base of the leaf petioles. The flowers of *C. mariana*, like those of many other plants in the pea family, have a showy upper petal (banner), two lateral petals (wings), and two lower petals (keel) that enclose the pistil and stamens. The floral pedicels are twisted, inverting the blooms so that they face upward and the banners are in the lowest position. Each flower is 5–6 cm long and the petals are pale blue to pink, with darker lines near the center of the banner. The fruits are linear, dehiscent pods 3–6 cm in length. (See Foerste, 1893, Britton and Brown 1913, Fernald 1950, Gleason and Cronquist 1991, Fantz 1993, Benda undated).



Left: Britton and Brown 1913, courtesy USDA NRCS 2024a. Right: Eric M. Powell, 2018.

Clitoria mariana is morphologically similar to *Centrosema virginianum*, a species that is currently ranked as historical in New Jersey. *Centrosema* flowers have a spur-like projection on the back of the banner (Fernald 1950). Vegetative plants might be confused but the upper surfaces of *C. virginianum* leaves are slightly sticky to the touch while those of *Clitoria mariana* are smooth (Weakley et al. 2024). Trelease (1879) observed that the stems of *Centrosema virginianum* were often considerably longer than those of *Clitoria*, sometimes ascending to heights of three meters or more.

Throughout its range, showy flowers may be seen on *Clitoria mariana* plants from May through September and smaller flowers that remain closed can be produced through November, with fruit developing between June and November (Barton 1818, Johnson 1888, Ortenburger 1928, Platt et al. 1988, Rhoads and Block 2007, Weakley et al. 2024). In New Jersey the Butterfly-pea usually flowers from mid-July to mid-August and fruits between mid-August and mid-September (Stone 1911, Hough 1983). Aboveground parts of *C. mariana* plants generally die back to the ground by winter, emerging again in the spring (Johnson 1997, Judge 2022). The plants store energy and water in swollen underground stems (tubers) that may be joined in a series and oriented either vertically or horizontally (Diaz-Toribio and Putz 2021). *C. mariana* can also develop spherical nodules on the roots: The nodules harbor motile bacteria that transform atmospheric nitrogen into a form that can be used by the plants. Shunk (1921) reported that the nitrogen-fixing bacteria in *C. mariana* nodules have a single flagellum, although many species in the Fabaceae have multi-flagellate nodule bacteria. In comparison to some other legumes that grow in a comparable habitat, *C. mariana* produces smaller and fewer nodules and accumulates nitrogen slowly but over a long period it can amass significant amounts (Cathey et al. 2010).

Pollinator Dynamics

Clitoria flowers attract pollinators with a striking visual display and reward them with nectar, which is produced at the base of the ovary (Zomlefer 1994). The banner of *C. mariana* serves as a landing platform for insects and the dark markings on that petal direct them toward the nectar supply (Trelease 1879, Fantz 1993). In an essay about springtime in Florida, Nehrling (1904) noted that the blooms of *Clitoria mariana* were "at times very sweet-scented" but he may have conflated *C. mariana* and *C. fragrans*—the latter is a highly fragrant species that is restricted to Florida and had not yet been described at the time of his writing (see Small 1926).

Clitoria species are primarily cross-pollinated by bees. The structure of *C. mariana* flowers forces the bees to push their way inside to access nectar and pollen is deposited on their bodies in the process. Butterflies have also been observed nectaring on *Clitoria* blooms but they can apparently do so without providing pollination services (Trelease 1879, Foerste 1893, Zomlefer 1994, Fidalgo et al. 2018, Campbell 2020). Female Pipevine Swallowtails (*Battus philenor*) often alight on *C. mariana* leaves while searching for the *Aristolochia* species that serve as their larval host plants (Rausher and Papaj 1983).

During his pollination studies, Trelease (1879) was unable to determine whether *Clitoria mariana* was self-compatible. The species is now known to produce cleistogamous flowers, which are self-fertilizing by definition because they never open. The cleistogamous flowers of *C. mariana* may have been overlooked by early botanists since they usually develop later in the season than the showy flowers (Benda undated, Weakley et al. 2024) and they are considerably smaller. The calyx tubes of cleistogamous *C. mariana* flowers are 4–5 mm long, in contrast with those of the chasmogamous (outcrossing) flowers which are 10–14 mm long (Fantz 1993). Self-fertility has also been documented in other *Clitoria* species (Fantz 1993, Fidalgo et al. 2018, Campbell 2020), and the majority of *C. fragrans* flowers in a population studied by Lewis (2007) were cleistogamous.

Seed Dispersal and Establishment

Clitoria mariana seeds are about 4 mm in diameter (Lackey 2007). They have been described as pea-like and brown with a sticky surface, and Leggett (1876) observed that they retained their sticky quality for years after drying. When *Clitoria* fruits become dry the segments (valves) twist and separate, ejecting the seeds (Lewis 2007, CPC 2020). The mechanism helps to remove the propagules from the immediate vicinity of the parent plants but they remain local.

When Clark (1940, 1941) first documented *C. mariana* in Arizona he noted that a long-distance dispersal mechanism must be in play because the plants were about a thousand miles from the nearest known populations. The distribution of *C. mariana* seeds over long distances is probably accomplished by animals. The sticky seed coats may help them adhere to fur or feathers (CPC 2020) but dispersal via the consumption and defecation of viable seeds is likely to occur with greater frequency. *Clitoria mariana* seeds are an important food source for Northern Bobwhites (*Colinus virginianus*), particularly during the summer, and they are utilized by an assortment of other birds as well (Davidson 1942, Jones and Chamberlain 2004, Allain and Reid undated). Mammals may also make a significant contribution to dispersal. *C. mariana* is a preferred forage plant for White-tailed Deer (*Odocoileus virginianus*) and its high protein content makes it particularly valuable to nursing does in the summer (Warren and Hurst 1981, Loman et al. 2017, Glow et al. 2019) at a time when some of the plants may already be in fruit. Mammalian dispersal has been documented in another species: For example, viable seeds of *Clitoria ternatea* were recovered from elephant dung (Samansiri and Weerakoon 2008).

It is not clear whether *Clitoria mariana* forms a seed bank. Seeds of the species were not detected in a number of South Carolina sites where it was present in the vegetation (Cushwa et al. 1970). *C. mariana* seeds contain little endosperm and no starch (Lackey 2007). They germinate at the soil surface, initially producing smooth cotyledons on a hairy stem and a slender primary root. The first true leaves are opposite and simple and as the plants continue to grow they develop a short rhizome and some lateral roots but the primary root persists (Holm 1891). *C. mariana* is probably capable of forming mycorrhizal associations. The majority of species in the Fabaceae are mycorrhizal, and it appears that a greater uptake of phosphorous—facilitated by mycorrhizae—can enhance the performance of nitrogen-fixing bacteria (Bethanfalfay 1992, Wang and Qiu 2006, Püschel et al. 2017).

Habitat

Clitoria mariana has been recorded in a diverse array of habitats ranging from coastal lowland sites (Negrete et al. 1999, Stalter et al. 1999 & 2018) to mountainous locations at elevations of more than 2,000 meters above sea level (Clark 1940). Typical substrates are sand or shale barrens (Stone 1911, Clark 1940 & 1941, Gurney 1941, Wood 1944, Kral 1955, Duke 1961, Carr 1965, Hough 1983, Platt et al. 1988, Stalter and Lamont 1997, Rhoads and Block 2007, Carter and Floyd 2013, Judge 2022), but it has also been found growing on red clay (Johnson 1888) and on traprock (NJNHP 2024). Moisture conditions may range from mesic to xeric (McVaugh 1943, Kral 1955, McCoy 1958, Duke 1961, Carr 1965, Harris and Chester 1999, Hoagland and Buthod 2006, Glow and Ditchkoff 2017).

Clitoria mariana can thrive under a variety of light regimes, doing nearly as well in shade as in open situations (Weakley et al. 2024). It is often associated with open woodlands or edges but has been documented in both fully exposed and shaded sites (Clark 1941, Wood 1944, McGregor and Horr 1950, McCoy 1958, Carr 1965, Hough 1983, Houle 1987, Rhoads and Block 2007, NJNHP 2024). Brown (1970) categorized *C. mariana* as a shade-type plant, meaning that it is most photosynthetically efficient in weak light but is capable of tolerating higher light intensities without injury.

Wooded sites where *Clitoria mariana* occurs are most likely to be dominated by oaks (*Quercus* spp.), pines (*Pinus* spp.) or a mixture of both (Deam 1934, Clark 1940, Oosting 1942, Wood 1944, Silberhorn 1970, Clinton et al. 1993, Harris and Chester 1999, Carter and Londo 2006, Philipps et al. 2007, Boyle et al. 2009, Cipollini et al. 2012, Carter and Floyd 2013, Palmquist et al. 2014, Glow and Ditchkoff 2017, NJNHP 2024). It may also grow in oak-hickory (*Quercus-Carya* spp.) forests (Kral 1955, Rodgers and Shake 1965, Clements and Wofford 1991, Hill et al. 2009) and woodlands dominated by *Liquidambar styraciflua* (Ortenburger 1928) or *Sabal* spp. (Lawrence 2008). Tompkins and Bridges (2013) noted that *C. mariana* was weakly associated with Carolina prairies, and Crandall and Tyrl (2006) indicated that it was common in Oklahoma grasslands where *Schizachyrium scoparium* and *Andropogon gerardii* were dominant. Disturbed sites utilized by *C. mariana* include managed or abandoned fields and pastures, pine plantations, and roadsides (Williamson 1909, McVaugh 1943, Kral 1955, Lawrence 2008, Palmer et al. 2012, Weakley et al. 2024).

Wetland Indicator Status

Clitoria mariana is a facultative upland species, meaning that it usually occurs in nonwetlands but may occur in wetlands (U. S. Army Corps of Engineers 2020).

USDA Plants Code (USDA, NRCS 2024b)

The code for *Clitoria mariana* is CLMA4. The USDA does not have a code for var. *mariana*.

Coefficient of Conservancy (Walz et al. 2020)

CoC = 7. Criteria for a value of 6 to 8: Native with a narrow range of ecological tolerances and typically associated with a stable community (Faber-Langendoen 2018).

Distribution and Range

The native range of *Clitoria mariana* includes parts of the central and eastern United States and northern Mexico (POWO 2024). The map in Figure 1 depicts the extent of the species in the United States and Canada.

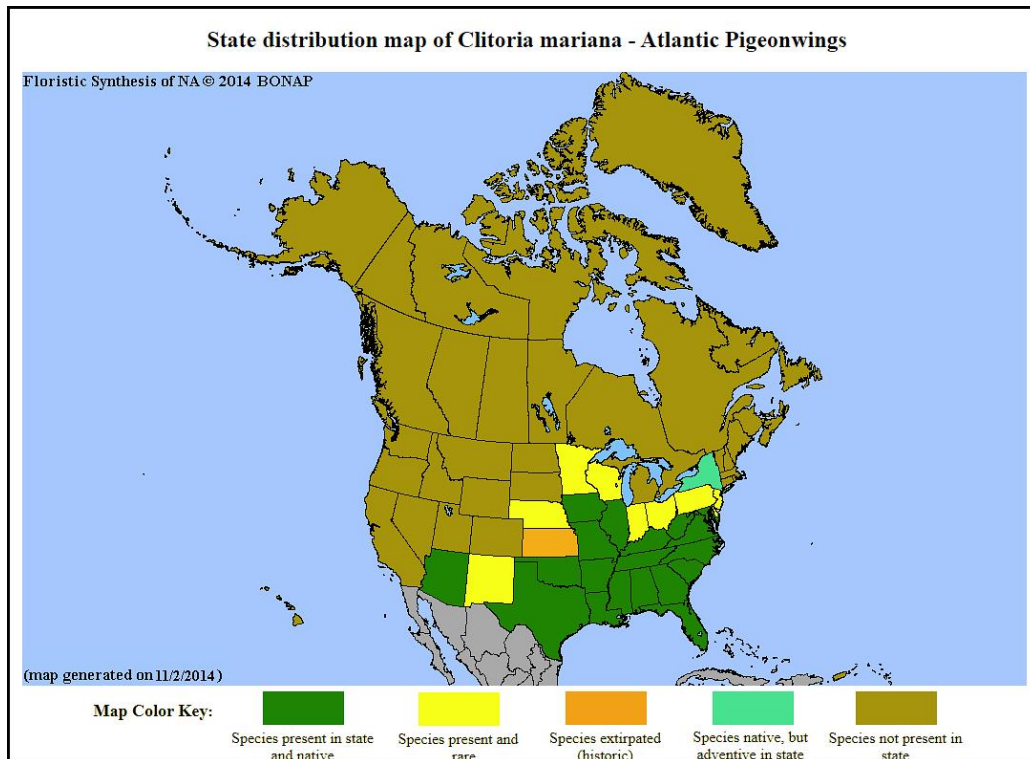


Figure 1. Distribution of *C. mariana* in the United States and Canada, adapted from BONAP (Kartesz 2015).

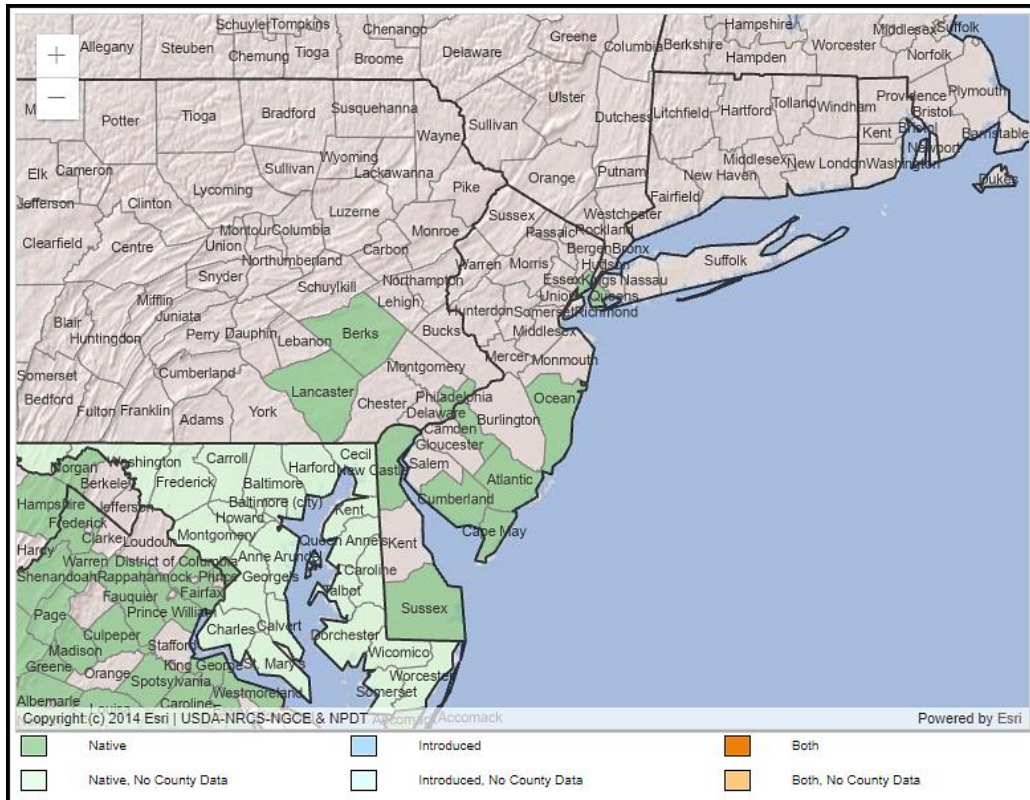


Figure 2. County records of *C. mariana* in New Jersey and vicinity (USDA NRCS 2024b).

The USDA PLANTS Database (2024b) shows records of *Clitoria mariana* in six New Jersey counties: Atlantic, Camden, Cape May, Cumberland, Hudson, and Ocean (Figure 2). The data include historic observations and do not reflect the current distribution of the species.

Conservation Status

Clitoria mariana is considered globally secure. The G5 rank means the species has a very low risk of extinction or collapse due to a very extensive range, abundant populations or occurrences, and little to no concern from declines or threats. The variety has not been ranked (NatureServe 2024). The map below (Figure 3) illustrates the conservation status of *C. mariana* in the United States. It is shown as vulnerable (moderate risk of extinction) in one state, imperiled (high risk of extinction) in two states, and critically imperiled (very high risk of extinction). *C. mariana* is not considered native in New York, although Fantz (2000) noted that the species was collected in that state during 1843. In most of the other states where it occurs Butterfly-pea is secure, apparently secure, or unranked. However in states like New Jersey where it is listed at the varietal level it may also appear to be unranked.

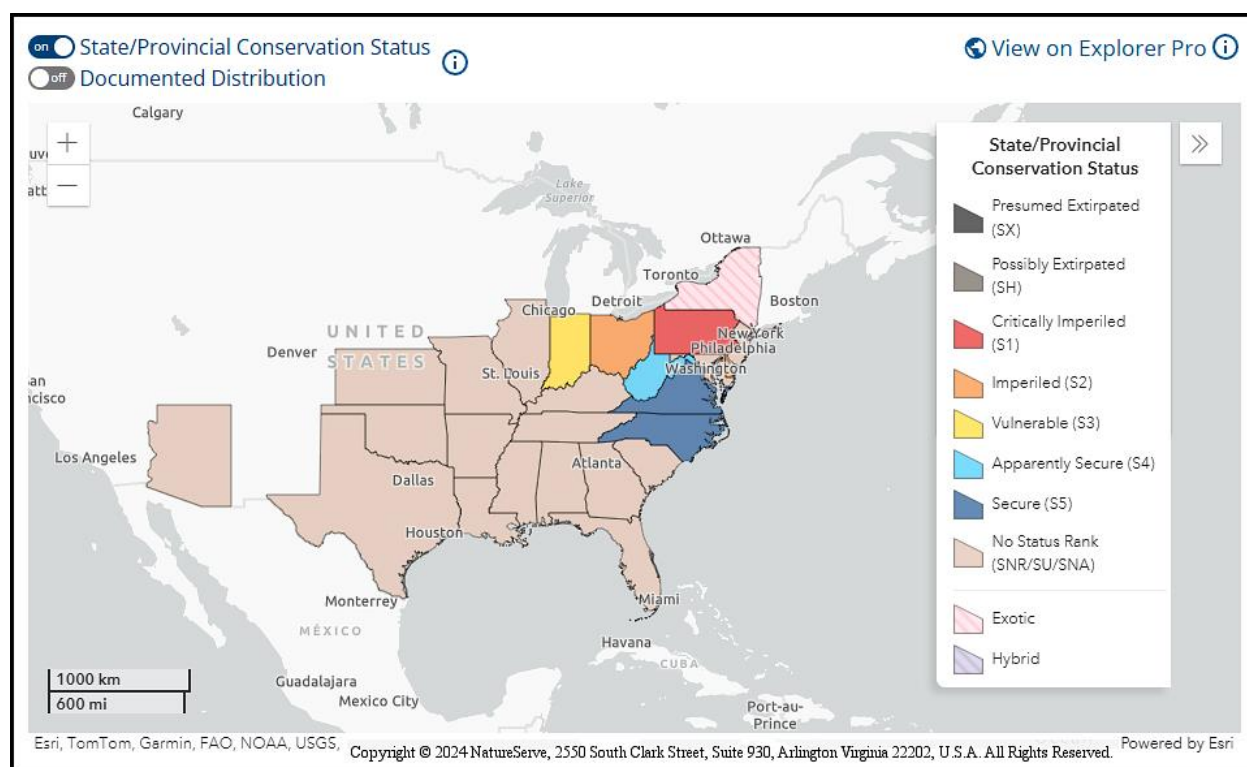


Figure 3. Conservation status of *C. mariana* in the United States (NatureServe 2024).

Clitoria mariana var. *mariana* is critically imperiled (S1) in New Jersey (NJNHP 2024). The rank signifies five or fewer occurrences in the state. A species with an S1 rank is typically either restricted to specialized habitats, geographically limited to a small area of the state, or significantly reduced in number from its previous status. *C. mariana* var. *mariana* is also listed as an endangered species (E) in New Jersey, meaning that without intervention it has a high likelihood of extinction in the state. Although the presence of endangered flora may restrict

development in certain communities such as wetlands or coastal habitats, being listed does not currently provide broad statewide protection for the plants. Additional regional status codes assigned to Butterfly-pea signify that the species is eligible for protection under the jurisdictions of the Highlands Preservation Area (HL) and the New Jersey Pinelands (LP) (NJNHP 2010).

Clitoria mariana was first documented in New Jersey by Asa Gray in 1833, and for nearly four decades it was only known from Ocean County (Stone 1911). In 1871 William Leggett collected the species at a site in Hudson County—the plants were in fruit at the time and the species was subsequently determined by Gray (Leggett 1876). At the time, the collector remarked "*There was a considerable number of plants in one spot. We trust that those who seek it there will gather it sparingly and help to preserve it from extermination.*" Unfortunately, that population was eventually extirpated (Breden et al. 2006). Additional observations of *C. mariana* were made in several of the southern counties, but the species was generally noted to be rare in the state (Willis 1974, Britton 1889, Keller and Brown 1905, Stone 1911, Taylor 1915). Its status was described as undetermined by Calazza and Fairbrothers (1980), and Hough (1983) noted that there was only a single current record. The population referenced by Hough was declining in 1990 and could not be relocated in 2007. Three additional occurrences were documented after her book was published but two of those have since been lost and just one is still extant (NJNHP 2024).

Threats

In New Jersey, documented losses of *Clitoria mariana* populations have been associated with both disturbances that directly damaged the plants (e.g. mowing, ditching, tire ruts, trail clearing) and the proliferation of invasive plants (NJNHP 2024). The spread of an invasive grass may also have contributed to the decline of a population in Mexico (Muñoz 2009).

While Butterfly-pea is able to grow in a variety of light conditions its vigor may decrease in heavily shaded sites. *Clitoria mariana* has been known to survive, or even benefit from, natural canopy disturbances caused by high winds or tornado activity, although not at sites where salvage harvesting followed a wind event (Brewer et al. 2012, Kleinman and Hart 2018). The species has often been associated with fire-prone habitats (Clinton et al. 1993) and is tolerant of management strategies used to control the growth of woody vegetation including prescribed burns, imazapyr-based herbicides, or a combination of both (Wigley et al. 2000, Jones and Chamberlain 2004, Hutchinson 2005, Iglay et al. 2010, Cipollini et al. 2012).

Although the overabundance of White-tailed Deer was not identified as a specific threat to *Clitoria mariana* populations in New Jersey, the detrimental impact on other native herbs in the state is well-documented (Kelly 2019). Butterfly-pea is preferentially browsed by deer throughout the growing season, but particularly during the spring and summer months (Warren and Hurst 1981, Iglay et al. 2010, Loman et al. 2017, Glow et al. 2019, Kroeger et al. 2020). The impact of deer herbivory could vary depending on the timing, intensity, or frequency. Grazing reportedly accelerated the growth of *Clitoria ternatea* (Campbell 2020), and—as previously discussed—late summer browsing on *C. mariana* might promote the long-distance dispersal of

its seeds. However, intense levels of herbivory early in the season could prevent fruit set or result in a loss of vigor.

Some insect herbivory has also been noted on *Clitoria mariana*: In particular a lace bug (*Leptopharsa clitoriae*), a larval butterfly (*Urbanus proteus*) and a larval moth (*Triclonella pergandeella*), all of which are specialist feeders on plants in the Fabaceae (Heidemann 1911, Skinner 1911, McAtee 1923, Tyler 2020). A leaf spot fungus, *Cercospora clitoriae*, was first described after it was found on the leaves of *Clitoria mariana* (Atkinson 1891). These pests and diseases can do some damage to *C. mariana* plants but there is no indication that they pose a significant threat to healthy populations.

Climate Change Vulnerability

Information from the references cited in this profile was used to evaluate the vulnerability of New Jersey's *Clitoria mariana* populations to climate change. The species was assigned a rank from NatureServe's Climate Change Vulnerability Index using the associated tool (Version 3.02) to estimate its exposure, sensitivity, and adaptive capacity to changing climatic conditions in accordance with the guidelines described by Young et al. (2016) and the state climatic computations by Ring et al. (2013). Based on available data *C. mariana* was assessed as Less Vulnerable, meaning that climate change is not expected to have a notable detrimental impact on its extent in New Jersey by 2050.

Some of the effects of changing climatic conditions in New Jersey include higher temperatures, a longer growing season, and shifting precipitation patterns that have increased the frequency and intensity of both droughts and floods (Hill et al. 2020). *Clitoria mariana* is near the northern end of its range in New Jersey so it will probably be able to adapt to warmer conditions, and as a species that can grow in xeric environments it is likely to have some capacity for drought tolerance. Rising temperatures may even give the species an opportunity to expand its range to the north, although the extent of its ability to withstand cold or freezing is unclear. However, *C. mariana* could face some additional threats from invasive plants, which are expected to become an even greater problem in the northeast as a result of climate change (Bellard et al. 2013, Salva and Bradley 2023).

Management Summary and Recommendations

In natural settings, *Clitoria mariana* benefits from prescribed fire. In Missouri Butterfly-pea is a characteristic plant of dry woodlands that have been burned repeatedly over an extended period (Stephen et al. 2024). Generally the species has been found to increase in cover at sites where it was already present (Taft 2003 & 2020, Cram et al. 2009), peaking in abundance two years after a fire (Buckner and Landers 1979). The most beneficial burns have usually been carried out during the winter or early spring while *C. mariana* was dormant, and intervals of 1–6 years appear to be effective (DeSelm et al. 1973, Sparks et al. 1998, Kush et al. 2000, Philipps et al. 2007, Glow 2016, Taft 2020). Late spring burns that are conducted after the plants have emerged may be detrimental (Cushwa et al. 1970).

According to Crouch (2010), *Clitoria mariana* may also respond positively to alternative management techniques for slowing succession such mowing, roller chopping, or disking but the timing of the events is likely to be important. Detrimental impacts from mowing have been observed at the site of New Jersey's only extant occurrence (NJNHP 2024). That population is not a candidate for management with fire because it is situated on the shoulder of a road and may also be on private property. In that situation, it would be more practical to identify the landowner and attempt to establish a cooperative agreement for protection of the rare plants. It is possible that additional populations could turn up in the state: The sites of a number of historical occurrences have never been searched and *C. mariana* is capable of utilizing a variety of habitats so it may have colonized some new locations.

Although there is a fair amount of information available about *Clitoria mariana* there are still a few outstanding questions regarding the species. For example, Johnson (2012) observed very low seed set in two western occurrences and the productivity of typical populations does not appear to have been studied. Additional topics suggested for further investigation include presence and persistence in the seed bank, cold tolerance, and the impacts of deer herbivory.

Synonyms

The accepted botanical name of the species is *Clitoria mariana* L. var. *mariana*. Orthographic variants, synonyms, and common names are listed below (ITIS 2024, POWO 2024, USDA NRCS 2024b). Some sources accept two additional varieties: *C. mariana* var. *pubescentia* is restricted to Florida and var. *orientalis* is endemic to southeast Asia (POWO 2024, Weakley et al. 2024). Other sources do not recognize any subtaxa (eg. Kartesz 2015, USDA NRCS 2024b). It is also worth noting that the common name 'Butterfly-pea' has been applied to more than one plant species.

Botanical Synonyms

Clitoria grahamii Steud.
Martiusia mariana (L.) Small

Common Names

Butterfly-pea
Atlantic Pigeonwings
She-pea

References

Allain, Larry and Chris Reid. Undated. *Clitoria mariana*. Species page on Plants of Louisiana, USGS Wetland and Aquatic Research Center. Accessed February 2, 2024 at <https://warcapps.usgs.gov/PlantID/Species/Details/720>

Atkinson, George F. 1891. Some Cercosporae from Alabama. Journal of the Elisha Mitchell Scientific Society 8(2): 33–67.

Barton, William P. C. 1818. Compendium Floræ Philadelphicæ: Containing a Description of the Indigenous and Naturalized Plants Within A Circuit of Ten Miles Around Philadelphia. Volume II. M. Carey and Son, Philadelphia, PA. 234 pp.

Bellard, C., W. Thuiller, B. Leroy, P. Genovesi, M. Bakkenes, and F. Courchamp. 2013. Will climate change promote future invasions? *Global Change Biology* 19(12): 3740–3748.

Benda, Christopher David. Undated. Butterfly Pea (*Clitoria mariana* L.) Plant of the Week - U. S. Forest Service, United States Department of Agriculture, Washington, D. C. Available at https://www.fs.usda.gov/wildflowers/plant-of-the-week/clitoria_mariana.shtml

Bethanfalvay, Gabor J. 1992. Mycorrhizal fungi in nitrogen-fixing legumes: Problems and prospects. *Methods in Microbiology* 24: 375–389.

Boyle, M. Forbes, Robert K. Peet, Thomas R. Wentworth, Michael P. Schafale, and Michael Lee. 2009. Natural Vegetation of the Carolinas: Classification and Description of Plant Communities of the Upper Tar, Roanoke, Meherrin, Chowan, and Cashie Rivers. Report prepared for the Ecosystem Enhancement Program, North Carolina Department of Environment and Natural Resources, Raleigh, NC. 67 pp.

Breden, T. F., J. M. Hartman, M. Anzelone and J. F. Kelly. 2006. Endangered Plant Species Populations in New Jersey: Health and Threats. New Jersey Department of Environmental Protection, Division of Parks and Forestry, Office of Natural Lands Management, Natural Heritage Program, Trenton, NJ. 198 pp.

Brewer, J. Stephen., Christine A. Bertz, Jeffery B. Cannon, Jason D. Chesser, and Erynn E. Maynard. 2012. Do natural disturbances or the forestry practices that follow them convert forests to early-successional communities? *Ecological Applications* 22(2): 442–458.

Britton, N. L. 1889. Catalogue of plants found in New Jersey. Geological Survey of New Jersey, Final Report of the State Geologist 2: 27–642.

Britton, N. L. and A. Brown. 1913. An Illustrated Flora of the Northern United States and Canada in three volumes: Volume II (Amaranth to Polypremum). Second Edition. Reissued (unabridged and unaltered) in 1970 by Dover Publications, New York, NY. 735 pp.

Brown, David Dee. 1970. Understory Light and Herbaceous Plant Photosynthesis in Oklahoma Forests. Doctoral Dissertation, University of Oklahoma, Norman, OK. 36 pp.

Buckner, James L. and J. Larry Landers. 1979. Fire and disking effects on herbaceous food plants and seed supplies. *The Journal of Wildlife Management* 43(3): 807–811.

Calazza, Nicholas and David E. Fairbrothers. 1980. Threatened and endangered vascular plant species of the New Jersey Pinelands and their habitats. Report prepared for the New Jersey Pinelands Commission. Available at <https://rucore.libraries.rutgers.edu/rutgers-lib/37776/PDF/1/play/>

- Campbell, Sean Michael. 2020. Germination, Growth, Flower Production and Quality of Butterfly Pea (*Clitoria ternatea*): A Plant With a Unique pH-dependent Sensory Quality. Doctoral Dissertation, University of Florida, Gainesville, FL. 151 pp.
- Carr, Lloyd G. K. 1965. Floristic elements in southwestern Virginia: A phytogeographical consideration. *Castanea* 30(2): 105–145.
- Carter, Robert and Robert Floyd. 2013. Landscape scale ecosystems of the Pine Mountain Range, Georgia. *Castanea* 78(4): 231–255.
- Carter, Robert and Andrew J. Londo. 2006. Remnant fire-disturbed montane Longleaf Pine forest in west central Georgia. *Proceedings of the 13th Biennial Southern Silvicultural Research Conference*, Memphis, Tennessee: 475–477.
- Cathey, Sarah E., Lindsay R. Boring, and Thomas R. Sinclair. 2010. Assessment of N₂ fixation capability of native legumes from the longleaf pine–wiregrass ecosystem. *Environmental and Experimental Botany* 67(3): 444–450.
- Cipollini, Martin L., Joshua Culberson, Cade Strippelhoff, Thomas Baldvins, and Kalia Miller. 2012. Herbaceous plants and grasses in a mountain Longleaf Pine forest undergoing restoration: A survey and comparative study. *Southeastern Naturalist* 11(4): 637–668.
- Clark, Ora M. 1940. A new station for *Clitoria mariana*. *Proceedings of the Oklahoma Academy of Science* 20: 63.
- Clark, Ora M. 1941. Other stations extend distribution of *Clitoria mariana*. *Proceedings of the Oklahoma Academy of Science* 21: 69.
- Clements, Richard K. and B. Eugene Wofford. 1991. The vascular flora of Wolf Cove, Franklin County, Tennessee. *Castanea* 56(4): 268–286.
- Clinton, B. D., J. M. Vose, and W. T. Swank. 1993. Site preparation burning to improve southern Appalachian pine-hardwood stands: Vegetation composition and diversity of 13-year-old stands. *Canadian Journal of Forest Research* 23: 2271–2277.
- CPC (Center for Plant Conservation). 2020. Sweetscented Pigeonwings (*Clitoria fragrans*). Accessed December 16, 2024 at <https://saveplants.org/plant-profile/?CPCNum=449614>
- Cram, Douglas S. Ronald E. Masters, Fred S. Guthery, David M. Engle, and Warren G. Montague. 2009. Usable space versus food quantity in Bobwhite habitat management. *National Quail Symposium Proceedings*, Volume 6: 146–159.
- Crandall, Raelene M. and Ronald J. Tyrl. 2006. Vascular flora of the Pushmataha Wildlife Management Area, Pushmataha County, Oklahoma. *Castanea* 71(1): 65–79.

Crouch, Tyson Lewis. 2010. Brood Habitat Use and Availability and Daily and Seasonal Covey Movements of Northern Bobwhites in East-central Alabama. Master's Thesis, Auburn University, Auburn, AL. 34 pp.

Cushwa, Charles T., Melvin Hopkins, and Burd S. McGinnes. 1970. Responses of legumes to prescribed burns in Loblolly Pine stands of the South Carolina piedmont. USDA Forest Service Research Note SE - 140, Southeastern Forest Experiment Station, Ashville, NC. 6 pp.

Davidson, Verne E. 1942. Bobwhite foods and conservation farming. The Journal of Wildlife Management 6(2): 97–109.

Deam, Chas. C. 1934. Plants new or rare to Indiana, XX. Proceedings of the Indiana Academy of Science 44: 53–54.

De Selm, H. R., E. E. C. Clebsch, G. M. Nichols, and E. Thor. 1973. Response of herbs, shrubs and tree sprouts in prescribed-burn hardwoods in Tennessee. Proceedings of the 13th Annual Tall Timbers Fire Ecology Conference. Tall Timbers Research Station, Tallahassee, FL: 331–344.

Diaz-Toribio, Milton H. and Francis E. Putz. 2021. Underground carbohydrate stores and storage organs in fire-maintained Longleaf Pine savannas in Florida, USA. American Journal of Botany 108(3): 432–442.

Duke, James A. 1961. The psammophytes of the Carolina fall-line sandhills. Journal of the Elisha Mitchell Scientific Society 77(1): 3–25.

Faber-Langendoen, D. 2018. Northeast Regional Floristic Quality Assessment Tools for Wetland Assessments. NatureServe, Arlington, VA. 52 pp.

Fantz, Paul R. 1993. Revising the classification of cultivated *Centrosema* and *Clitoria* in the United States. HortScience 28(6): 674–676.

Fantz, Paul R. 2000. Nomenclatural notes on the genus *Clitoria* for the Flora North American Project. Castanea 65(2): 89–92.

Fernald, M. L. 1950. Gray's Manual of Botany. Dioscorides Press, Portland, OR. 1632 pp.

Fidalgo, Adriana de Oliveira, Débora Marcouizos Guimarães, Gabriela Toledo Caldiron, and José Marcos Barbosa. 2018. Reproductive ecology of two pioneer legumes in a coastal plain degraded by sand mining. Hoehnea 45(1): 93–102.

Foerste, August F. 1893. Botanical notes from Bainbridge, Georgia. I. The Botanical Gazette 18(12): 456–466.

Gleason, H. A. and A. Cronquist. 1991. Manual of Vascular Plants of Northeastern United States and Adjacent Canada. Second Edition. The New York Botanical Garden, Bronx, NY. 910 pp.

Glow, Michael P. 2016. Prescribed Fire Interval and Economic Tradeoffs on Forage and Nutrient Availability During Stress Periods for White-tailed Deer. Master's Thesis, Auburn University, Auburn, AL. 110 pp.

Glow, Michael P. and Stephen S. Ditchkoff. 2017. Economic optimization of forage and nutrient availability during stress periods for White-tailed Deer. Journal of the Southeastern Association of Fish and Wildlife Agencies 4: 121–129.

Glow, Michael P., Stephen S. Ditchkoff, and Mark D. Smith. 2019. Annual fire return interval influences nutritional carrying capacity of White-Tailed Deer in pine–hardwood forests. Forest Science 65(4): 483–491.

Gurney, Ashley B. 1941. Taxonomic and bionomic notes on the grasshopper *Melanoplus impudicus* Scudder (Orthoptera: Acrididae). The American Midland Naturalist 26(3): 558–569.

Harris, Christine E. and Edward W. Chester. 1999. The herbaceous flora of mature Chestnut Oak forests, Northwestern Highland Rim, Kentucky and Tennessee. Journal of the Tennessee Academy of Science 74(1-2): 41–47.

Heidemann, O. 1911. A new species of North American Tingitidae. Proceedings of the Entomological Society of Washington 13: 180–182.

Hill, JoVonn, G., W. Douglas Stone, and Jennifer L. Seltzer. 2009. Description of an upland oak-hickory forest in the Black Belt from Osborn Prairie, Oktibbeha County, Mississippi. Journal of the Mississippi Academy of Sciences 54(2): 157–163.

Hill, Rebecca, Megan M. Rutkowski, Lori A. Lester, Heather Genievich, and Nicholas A. Procopio (eds.). 2020. New Jersey Scientific Report on Climate Change, Version 1.0. New Jersey Department of Environmental Protection, Trenton, NJ. 184 pp.

Hoagland, Bruce W. and Amy Buthod. 2006. The vascular flora of the J. T. Nickel Family Nature and Wildlife Preserve, Cherokee County, Oklahoma. Castanea 73(1): 16–28.

Holm, Theodore. 1891. Contributions to the knowledge of the germination of some North American plants. Memoirs of the Torrey Botanical Club 2(3): 57–108.

Hough, Mary Y. 1983. New Jersey Wild Plants. Harmony Press, Harmony, NJ. 414 pp.

Houle, Gilles. 1987. Vascular plants of Arabia Mountain, Georgia. Bulletin of the Torrey Botanical Club 114(4): 412–418.

Hutchinson, Todd. 2005. Fire and the herbaceous layer of eastern oak forests. In Fire in Eastern Oak Forests: Delivering Science to Land Managers. Proceedings of a Conference. USDA Forest Service Northern Research Station, General Technical Report NRS-P-1: 136–149.

Igley, Raymond B., Phillip D. Jones, Darren A. Miller, Stephen Demarais, Bruce D. Leopold, and L. Wes Burger Jr. 2010. Deer carrying capacity in mid-rotation pine plantations of Mississippi. *Journal of Wildlife Management* 74(5): 1003–1012.

ITIS (Integrated Taxonomic Information System). Accessed December 10, 2024 at <http://www.itis.gov>

Johnson, L. N. 1888. A tramp in the North Carolina mountains. *Botanical Gazette* 13(10): 269–271.

Johnson, M. B. 1997. Cold tolerance of legumes: Effects of the December 1996 freeze on DELEPS's Tucson fields. *Aridus* 9(3): 1–4, 8.

Johnson, Matthew B. 2012. Autumn in the season for seeds - DELEP/BTA collecting trips in 2012. *Desert Plants* 28(2): 17–22.

Jones, Judy D. J. and Michael J. Chamberlain. 2004. Efficacy of herbicides and fire to improve vegetative conditions for Northern Bobwhites in mature pine forests. *Wildlife Society Bulletin* 32(4): 1077–1084.

Judge, Phoebe Alexis. 2022. The Impacts of Hydraulic Redistribution on the Physiology and Growth of Understory Plants in Longleaf Pine (*Pinus palustris*) Sandhills. Master's Thesis, University of Georgia, Athens, GA. 48 pp.

Kartesz, J. T. 2015. The Biota of North America Program (BONAP). Taxonomic Data Center. (<http://www.bonap.net/tdc>). Chapel Hill, NC. [Maps generated from Kartesz, J. T. 2015. Floristic Synthesis of North America, Version 1.0. Biota of North America Program (BONAP) (in press)].

Keller, Ida A. and Stewardson Brown. 1905. Handbook of the Flora of Philadelphia and Vicinity. Philadelphia Botanical Club, Philadelphia, PA. 360 pp.

Kelly, Jay F. 2019. Regional changes to forest understories since the mid-Twentieth Century: Effects of overabundant deer and other factors in northern New Jersey. *Forest Ecology and Management* 444: 151–162.

Kleinman, Jonathan S. and Justin L. Hart. 2018. Vascular flora of Longleaf Pine woodlands after wind disturbance and salvage harvesting in the Alabama Fall Line Hills. *Castanea* 83(2): 183–195.

Kral, Robert. 1955. A floristic comparison of two hillside bog localities in northeastern Texas. *Field & Laboratory* 23(3&4): 47–69.

Kroeger, Anthony J., Christopher E. Moorman, Marcus A. Lashley, M. Colter Chitwood, Craig A. Harper, and Christopher S. DePerno. 2020. White-tailed deer use of overstory hardwoods in longleaf pine woodlands. *Forest Ecology and Management* 464: 118046.

Kush, John S., Ralph S. Meldahl, and W. D. Boyer. 2000. Understory plant community response to season of burn in natural Longleaf Pine forests. *In* W. Keith Moser and Cynthia E Moser (eds.). *Fire and Forest Ecology: Innovative Silviculture and Vegetation Management*. Tall Timbers Fire Ecology Conference Proceedings No. 21, Tall Timbers Research Station, Tallahassee, FL.

Lackey, James A. 2007. Cotyledon areoles in the Fabaceae subtribe Clitoriinae. *Journal of the Botanical Research Institute of Texas* 1(2): 1101–1118.

Lawrence, Mario James. 2008. Abundance, Richness, and Habitat Preferences of Small Mammals on Spring Island, South Carolina. Master's Thesis, Coastal Carolina University, Conway, SC. 42 pp.

Leggett, William H. 1876. *Clitoria mariana*. *Bulletin of the Torrey Botanical Club* 6(15): 84.

Lewis, Michelle Nicole. 2007. Life History and Reproductive Biology of *Clitoria fragrans* Relative to Fire History on the Avon Park Air Force Range. Master's Thesis, University of Central Florida, Orlando, FL. 44 pp.

Loman, Zachary G., Ethan J. Greene, Bradley R. Wheat, Stephen Demarais, Darren A. Miller, Scott A. Rush, and Samuel K. Riffell. 2017. White-tailed Deer carrying capacity, intercropping switchgrass, and pine plantations. *The Journal of Wildlife Management* 81(6): 999–1008.

McAtee, W. L. 1923. Tingitoidea of the vicinity of Washington, D. C. (Heteroptera). *Proceedings of the Entomological Society of Washington* 25(7-8): 143–151.

McCoy, Doyle Allen. 1958. Vascular plants of Pontotoc County, Oklahoma. *The American Midland Naturalist* 59(2): 371–396.

McGregor, R. L. and W. H. Horr. 1950. Kansas plants new to Kansas herbaria V. *Transactions of the Kansas Academy of Science* 53(3): 365–369.

McVaugh, Rogers. 1943. The vegetation of the granitic flat-rocks of the southeastern United States. *Ecological Monographs* 13(2): 119–166.

Metcalf, Kevin. 2021. Photo of *Clitoria mariana* from North Carolina. Shared via iNaturalist at <https://www.inaturalist.org/observations/91700493>, licensed by <https://creativecommons.org/licenses/by-nc/4.0/>

Muñoz, Alfonso de Jesus Sanchez. 2009. Invasive Lehmann Lovegrass (*Eragrostis lehmanniana*) in Chihuahua, Mexico: Consequences of Invasion. Doctoral Dissertation, Oklahoma State University, Stillwater, OK. 97 pp.

NatureServe. 2024. NatureServe Explorer [web application]. NatureServe, Arlington, VA. Accessed December 10, 2024 at <https://explorer.natureserve.org/>

Negrete, I. G., A. D. Nelson, J. R. Goetze, L. Macke, T. Wilburn, and A. Day. 1999. A checklist for the vascular plants of Padre Island National Seashore. SIDA, Contributions to Botany 18(4): 1227–1245.

Nehrling, H. 1904. The beginning of spring in Florida - II. The Plant World 7(5): 118–122.

NJNHP (New Jersey Natural Heritage Program). 2010. Explanation of Codes Used in Natural Heritage Reports. Updated March 2010. Available at https://nj.gov/dep/parksandforests/natural/docs/nhpcodes_2010.pdf

NJNHP (New Jersey Natural Heritage Program). 2024. Biotics 5 Database. NatureServe, Arlington, VA. Accessed March 15, 2024.

Oosting, Henry J. 1942. An ecological analysis of the plant communities of Piedmont, North Carolina. The American Midland Naturalist 28(1): 1–126.

Ortenburger, A. I. 1928. Plant collections representative of some typical communities of eastern Oklahoma. Proceedings of the Oklahoma Academy of Science 8: 53–57.

Palmer, W. E., D. C. Sisson, S. D. Wellendorf, A. M. Bostick III, T. M. Terhune, and T. L. Crouch. 2012. Habitat selection by Northern Bobwhite broods in pine savanna ecosystems. Proceedings of the National Quail Symposium 7: 108–112.

Palmquist, Kyle A., Robert K. Peet, and Susan C. Carr. 2014. Xeric Longleaf Pine Vegetation of the Atlantic and East Gulf Coast Coastal Plain: An Evaluation and Revision of Associations within the U.S. National Vegetation Classification. 38 pp.

Philipps, Thomas C., Suzanne Birmingham Walker, Barbara R. MacRoberts, and Michael H. MacRoberts. 2007. Vascular flora of a Longleaf Pine upland in Sabine County, Texas. Phytologia 89(3): 317–338.

Platt, William J., Gregory W. Evans, and Mary M. Davis. 1988. Effects of fire season on flowering of forbs and shrubs in Longleaf Pine forests. Oecologia 76: 353–363.

Powell, Eric M. 2018. Photo of *Clitoria mariana* from Florida. Shared via iNaturalist at <https://www.inaturalist.org/observations/37826442>, licensed by <https://creativecommons.org/licenses/by-nc/4.0/>

POWO. 2024. Plants of the World Online. Facilitated by the Royal Botanic Gardens, Kew. Accessed December 10, 2024 at <http://www.plantsoftheworldonline.org/>

Püschel, David, Martina Janoušková, Alena Voříšková, Hana Gryndlerová, Miroslav Vosátka, and Jan Jansa. 2017. Arbuscular mycorrhiza stimulates biological nitrogen fixation in two

Medicago spp. through improved phosphorus acquisition. *Frontiers in Plant Science* 8: Article 390.

Rausher, Mark D. and Daniel R. Papaj. 1983. Host plant selection by *Battus philenor* butterflies: Evidence for individual differences in foraging behavior. *Animal Behavior* 31: 341–347.

Rhoads, Ann Fowler and Timothy A. Block. 2007. *The Plants of Pennsylvania*. University of Pennsylvania Press, Philadelphia, PA. 1042 pp.

Ring, Richard M., Elizabeth A. Spencer, and Kathleen Strakosch Walz. 2013. Vulnerability of 70 Plant Species of Greatest Conservation Need to Climate Change in New Jersey. New York Natural Heritage Program, Albany, NY and New Jersey Natural Heritage Program, Department of Environmental Protection, Office of Natural Lands Management, Trenton, NJ, for NatureServe #DDCF-0F-001a, Arlington, VA. 38 pp.

Rodgers, C. Leland and Roy E. Shake. 1965. Survey of vascular plants in Bearcamp Creek Watershed. *Castanea* 30(3): 149–166.

Salva, Justin D. and Bethany A. Bradley. 2023. High-impact invasive plants expanding into mid-Atlantic states: Identifying priority range-shifting species for monitoring in light of climate change. *Invasive Plant Science and Management* 16: 197–206.

Samansiri, K. A. P and Devaka K. Weerakoon. 2008. A study on the seed dispersal capability of Asian elephants in the northwestern region of Sri Lanka. *Gajah* 28: 19–24.

Shunk, Ivan V. 1921. Notes on the flagellation of the nodule bacteria of Leguminosae. *Journal of Bacteriology*, 6(2): 239–247.

Silberhorn, Gene M. 1970. A distinct phytogeographic area in Ohio: The southeastern Allegheny Plateau. *Castanea* 35(4): 277–292.

Skinner, Henry. 1911. The larger boreal American Hesperidæ, including *Eudamus*, *Erycides*, *Pyrrhopyge* and *Megathymus*. *Transactions of the American Entomological Society* 37(3): 169–209.

Small, John K. 1926. A new butterfly-pea from Florida. *Torreyana* 26: 56–57.

Sparks, Jeffrey C., Ronald E. Masters, David M. Engle, Michael W. Palmer, and George A. Bukenhofer. 1998. Effects of late growing-season and late dormant-season prescribed fire on herbaceous vegetation in restored pine-grassland communities. *Journal of Vegetation Science* 9: 133–142.

Stalter, Richard and Eric E. Lamont. 1997. Flora of North Carolina's Outer Banks, Ocracoke Island to Virginia. *The Journal of the Torrey Botanical Society* 124(1): 71–88.

Stalter, Richard, Maritess Leyva, and Dwight T. Kincaid. 1999. The flora of Indian shell rings from coastal South Carolina to northern Florida. *SIDA, Contributions to Botany* 18(3): 861–875.

Stalter, Richard, John Baden, Chester DePratter, and Paul Kenny. 2018. The vascular flora of coastal Indian clam shell middens in South Carolina, U.S.A. *Journal of the Botanical Research Institute of Texas* 12(2): 697–706.

Stephen, Carolyn A., Dan G. Drees, Jamie H. Ladner, and Lauren L. Sullivan. 2024. Fire effects on plant communities in Ozark woodlands and glades. Preprint, published online January 8, 2024: <https://assets.researchsquare.com/files/rs-3771313/v1/3d688b01-f028-44e8-83f6-039d34363e24.pdf?c=1704743131>

Stone, Witmer. 1911. *The Plants of Southern New Jersey*. Quarterman Publications, Boston, MA. 828 pp.

Taft, John B. 2003. Fire effects on community structure, composition, and diversity in a dry sandstone barrens. *Journal of the Torrey Botanical Society* 130(3): 170–192.

Taft, John B. 2020. Do early trends in oak barrens fire treatment predict later outcomes? Insights from three decades of vegetation monitoring. *Fire Ecology* 16: Article 23.

Taylor, Norman. 1915. Flora of the vicinity of New York - A contribution to plant geography. *Memoirs of the New York Botanical Garden* 5: 1–683.

Tompkins, Robert D. and William C. Bridges Jr. 2013. Restoration and plant species diversity of an Eastern prairie. *Native Plants* 14(2): 101–113.

Trelease, William. 1879. The fertilization of our native species of *Clitoria* and *Centrosema*. *The American Naturalist* 13(11): 688–692.

Tyler, Royal. 2020. Notes on the life history and food plants of *Triclonella pergandeella* Busck. *Southern Lepidopterists' News* 42(4): 313–318.

U. S. Army Corps of Engineers. 2020. National Wetland Plant List, version 3.5. https://cwbi-app.sec.usace.army.mil/nwpl_static/v34/home/home.html U. S. Army Corps of Engineers Research and Development Center, Cold Regions Research and Engineering Laboratory, Hanover, NH.

USDA, NRCS (U. S. Dept. of Agriculture, Natural Resources Conservation Service). 2024a. *Clitoria mariana* illustration from Britton, N. L. and A. Brown, 1913, *An illustrated flora of the northern United States, Canada and the British Possessions*, 3 vols., Kentucky Native Plant Society, New York, Scanned By Omnitek Inc. Image courtesy of The PLANTS Database (<http://plants.usda.gov>). National Plant Data Team, Greensboro, NC.

USDA, NRCS (U. S. Dept. of Agriculture, Natural Resources Conservation Service). 2024b. PLANTS profile for *Clitoria mariana* (Atlantic Pigeonwings). The PLANTS Database, National Plant Data Team, Greensboro, NC. Accessed December 10, 2024 at <http://plants.usda.gov>

Walz, Kathleen S., Jason L. Hafstad, Linda Kelly, and Karl Anderson. 2020. Floristic Quality Assessment Index for Vascular Plants of New Jersey: Coefficient of Conservancy (CoC) Values for Species and Genera (update to 2017 list). New Jersey Department of Environmental Protection, New Jersey Forest Service, Office of Natural Lands Management, Trenton, NJ.

Wang, B., and Y. L. Qiu. 2006. Phylogenetic distribution and evolution of mycorrhizas in land plants. *Mycorrhiza* 16(5): 299–363.

Warren, R. C. and G. A. Hurst. 1981. Rating of Plants in Pine Plantations as White-tailed Deer Food. Mississippi Agricultural & Forestry Experiment Station Information Bulletin 18, Mississippi State, MS. 14 pp.

Weakley, A. S. and Southeastern Flora Team. 2024. Flora of the Southeastern United States. Edition of March 4, 2024. University of North Carolina Herbarium, North Carolina Botanical Garden, Chapel Hill, NC. 2023 pp.

Wigley, T. Bentley, Karl V. Miller, David S. deCalesta, and Mark W. Thomas. 2000. Herbicides as an alternative to prescribed burning for achieving wildlife management objectives. In The Role of Fire in Nongame Wildlife Management and Community Restoration: Traditional Uses and New Directions. Proceedings of a Special Workshop. USDA Forest Service Northern Research Station General Technical Report NE-288: 124–138.

Williamson, Charles S. 1909. Notes on the flora of central and southern Delaware. *Torreyia* 9(8): 160–166.

Willis, O. 1874. Catalogue of Plants Growing in the State of New Jersey. J. W. Schermerhorn, New York, NY. 92 pp.

Wood, Carroll E. Jr. 1944. Notes on the flora of Roanoke County, Virginia (continued). *Rhodora* 46(544): 135–142.

Young, Bruce E., Elizabeth Byers, Geoff Hammerson, Anne Frances, Leah Oliver, and Amanda Treher. 2016. Guidelines for Using the NatureServe Climate Change Vulnerability Index, Release 3.02, 1 June 2016. NatureServe, Arlington, VA. 65 pp.

Zomlefer, Wendy B. 1994. Guide to Flowering Plant Families. University of North Carolina Press, Chapel Hill, North Carolina. 430 pp.